

# Journal

# SURVEY AND IDENTIFICATION OF MAJOR FUNGI CAUSING ROOT ROT ON DATE PALM AND THEIR RELATIVE IMPORTANCE IN EGYPT

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#### **ABSTRACT**

Date palms under the Egyptian conditions are subjected to infection with different diseases caused by many soil-borne pathogenic fungi which may cause considerable losses in the offshoots and trees. A survey was carried out during four years 2005-2008 in seven governorates. Aswan governorate showed the highest disease severity (45.00%), followed by Luxor (37.50%), Behaera (30.50%), Marsa-Matrouh (25.00%), Ismailia (5.00%), Sharkyia (3.75%) and Giza (2.50%). Isolation and identification of associated fungi showed that the most frequent fungi were Fusarium spp. and Thielaviopsis paradoxa, while the least frequent were Botrydiplodia theomromae and Rhizctonia solani. The most virulent fungi were F. oxysporum, F. moniliforme, F. solani and T. paradoxa which were responsible for the root rot incidence that started with gradual yellowing that reached the palm tip followed by quick death. All the tested cultivars were susceptible to infection by the isolated pathogenic fungi. Hayany cultivar was the most susceptible to infection, followed by Sammany cultivar. While Zaghloul cultivar was the least susceptible.

**Key Words:** Date palm, root rot, soil borne pathogenic fungi, Fusarium oxysporum, F. moniliforme, F. solani, Thielaviopsis paradoxa, Botrydiplodia theomromae, Rhizctonia solani

### INTRODUCTION

Date palm (*Phoenix dactylifera* L.) roots are liable to attack by several pathogenic soil borne fungi, that causing serious diseases. Plant pathogens, viz. fungi, nematodes, bacteria, and viruses can cause diseases or damages in palm trees (Chase and Broschat., 1991). Several fungi were recorded as causal pathogens of root rot on date palm, viz., Fusarium oxysporum; F. solani; F. moniliforme; F. semitectium; F. equiseti; Phoma sp.; Chaetomium sp.; Alternaria sp.; Cladosporium sp.: Macrophomina phaseolina; *Thielaviopsis* paradoxa: Diplodia phoenicum; Phomopsis phoenicola Rhizoctonia solani (Abdalla et al., 2000; El- Zawahry et al., 2000; Sarhan, 2001; Suleman, et al., 2001; El-Morsi, 2004; Mansoori and Kord, 2006; El Deeb et al., 2007; Samir et al., 2009). This study aimed to throw some light on the distribution of date palm root rot and its causal agents in different governorates in Egypt.

# MATERIALS AND METHODS

# Field survey:

Roots and soil (rhizosphere) samples were collected from naturally infected date palm and offshoots growing in different locations in seven governorates, *viz.*; Aswan, Luxor, Marsa-Matrouh, Giza, Ismailia, Sharkyia and Behara. These were selected depending on, the large area cultivated, irrigation system, environmental conditions (during different growing seasons, summer, autumen, winter and spring, 2005-2008) and type of soils (Sarhan, 2001 and Suleman, 2001). Disease incidence and Disease severity Index (DSI) were carried out according to Cooke *et al.* (2006) and the scale was suggested by Abdalla *et al.* (2000).

#### Isolation and identification:

Date palm roots and rhizoshpere (trees or offshoots) were collected from orchard infested localities and carefully clarified from the large particles of soil. Samples of roots were taken at least 15-20 cm apart, then maintained in a cool dry state during transit to the laboratory to minimize contamination with the saprophytic microorganisms. Samples were washed carefully with tap water to remove the adhering soil particles, and ten pieces (10 cm length) representing each sample were sectioned into small pieces (1 cm length) and

surface sterilized with 1% sodium hypochlorite solution for 2 mins. washed in sterile distilled water (SDW), and then dried between folds of sterilized filter papers. The sterilized root sections were transferred on potato dextrose agar (PDA) and the plates were incubated at 25±2°C. Emerged fungi were isolated and purified using the single spore technique and/or the hyphal tip method according to Wang and Wen (1997). The frequency of the isolated fungi was calculated separately from each collected sample. Stock cultures were maintained on PDA slants and kept in a refrigerator at 5°C, for further studies. Stocks were routinely sub-cultured on fresh slant every month. The fungal colonies growing in the culture plates were identified according to their morphological characteristics according to Nelson et al. (1983); Barnett and Hunter (1999); John and Summerell (2006). Micro-organisms from root surfaces (rhizosphere) were isolated according to Bao et al. (2004). The frequency of the isolated fungi from the root rotted samples and from rhizosphere was separately calculated according to the following formula: % Fungal frequency= Number of isolates of each fungs/Total number of all isolates X 100

# Pathogenicity test of the most frequent isolated fungi:

Pathogenicity test of the most frequent fungi, i.e. Fusarium oxysproum Schlecht, Fusarium solani (Mort.) Sacc, Fusarium moniliforme Sheldon, Fusarium semitectum Berk and Ravenel. Botryodiplodia theobrome Pat, Thielaviopsis thielavioides Peyr. and Rhizoctonia solani Kuhn., which were isolated from diseased roots was carried out in the greenhouse of Fruit and Woody Trees Diseases Research Department-Agric. Res. Center, Giza-Egypt. Seeds of Phoenix dactylifera were provided by a commercial producer. They were surface-disinfested for 10 min in a sodium hypochlorite solution NaOCl (1.5% available chlorine), soaked under tap water for 24 h, and then sown in black plastic bags (15 cm) filled with a sterilized mixture of equal portions (v/v) of soil, sand and clay. The seedlings were allowed to grow for 6 months or to the 2-3 leaves-stage. Five bags (each contained one date palm) representing each of the tested varities, viz. Zaghloul, Sammany and Hayany were used as rreplicates for each tested fungus. Seedlings and soil infested protocol was carried out by two methods:

**a- Seedlings injection:** Six-month-old plants were inoculated as described by Abdalla *et al.* (2000) by injecting 1 ml of hyphal or spore suspension, and the concentration was adjusted to 1X10<sup>6</sup>/ ml using a haemocytometer (Mather and Roberts, 1998). The hyphal and spore suspension were injected into the crown using a hypodermic needle and syringe for each fungus. After inoculation, all plants were covered separately with plastic bags for 48 h to maintain high humidity. Five plants of each cultivar were inoculated with each isolate, and corresponding controls were injected with SDW. The pots were arranged in a complete randomized design. The pathogenicity test was conducted twice. Pathogenicity was evaluated at 30, 45, 60 and 90 days after inoculation and disease reaction was rated as described before.

**b- Soil infestation:** Soil was infested according to El-Zawahry *et al.* (2000) by adding 100 ml/hyphal or spore suspension (4X10<sup>6</sup>/ml) to each black plastic bag representing each pathogenic fungus. Soil was irrigated every 3-4 days to ensure distribution of the tested fungus. After three months from soil infestation plants were uprooted and their roots were washed by water to remove soil particles, then percentage of infection and disease severity were recorded. Re-isolation was carried out from infected tissues.

#### Varietal reaction:

Reaction of the different varieties of date palm (Zaghloul, Sammany and Hayany) to infection with the different pathogenic fungi, viz. F. oxysproum, F. solani, F. monliforme, T. paradoxa, B. theobromae and R. solani was studied by using healthy 3-years-old offshoots. Soil infestation was adopted by adding 1000 ml/hyphal or spore suspension (4X10<sup>6</sup>/ ml) to each pot from each of the pathogenic fungi from active culture. In the other method for infection, plants were inoculated by injecting 1 ml of hypal or spore suspension (1X10<sup>6</sup>) of the tested fungi into the crown using a hypodermic needle and syringe. After inoculation, all plants were covered separately with plastic bags for 48 h to maintain high humidity. Five offshoots representing each variety were inoculated with each isolate, and corresponding controls were injected with sterilized distilled water (SDW). The Pathogenicity test was conducted twice. Pathogenicity was evaluated at 3, 6 and 9 month(s) after inoculation and disease reaction was rated as described before. The data were displayed in means after analysis of the least significant difference at 95% (LSD≤0.05) by Co-Stat Program (version 8.0).

#### RESULTS AND DISCUSSION

#### **Results**

#### **Field survey:**

Disease survey carried out during 2005-2008 growing seasons show clearly that typical symptoms of date palm root rot was observed in all governorates under study. Percentages of disease incidence as well as disease severity were found to be different from governorate to another, even from cultivar to the other. Data, (Table, 1) and (Fig. 1) show the disease incidence and disease severity percentages of root rot affected all studied cultivars of date palm in the different inspected locations in all governorates. The percentage of disease incidence and disease severity values were differed by the locality. The highest percentage of disease incidence (DI%) and severity (DS%) of root rot were recorded in Aswan (80%DI and 45%DS) followed by Luxor, Behera and Marsa-Matrouh (65%DI and 37.50%DS), (60%DI and 30%DS) and (50%DI and 25%DS), respectively, whereas the lowest percentage of disease incidence and severity were observed in Ismailia, Sharkvia and Giza (20%DI and 5%DS), (15%DI and 3.75%DS) and (10% DI and 2.5%DS), respectively.

Table (1): Field survey of root rot of different cultivars of date palm in different governorates.

Governorates	Locations	Total number of date palm trees*	Disease incidence%	Disease Severity
Marsa-Matrouh	Siwa	436344	50.00	25.00
Behera	Rashid – Edko	1128466	60.00	30.00
Sharkyia	Belbeas	1214798	15.00	3.75
Ismailia	Al- Ferdan	479766	20.00	5.00
Giza	El-Mansoria	514586	10.00	2.50
Luxor	Luxor	38393	65.00	37.50
Aswan	Abo El Ryesh- Al Akab	1008429	80.00	45.00

<sup>\*</sup>Anonymous (2009). Study of Important Indicators of the Agricultural Statistics. Ministry of Agriculture and Soil Reclamation. Economic Affairs Section, Egypt.

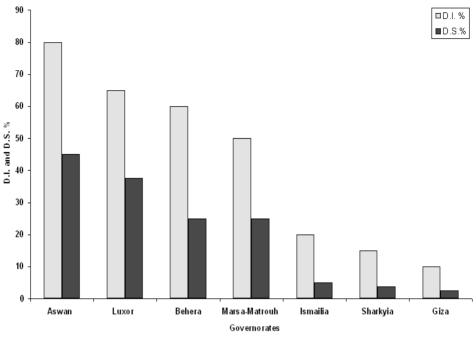


Fig. (1): Disease incidence and disease severity percentage of root rot of date palm in different governorates.

#### Isolation and identification:

Eight fungi, viz. Fusarium sp., Botryodiplodia theobromae, Thielaviopsis paradoxa, Gliocladium sp., Rhizoctonia solani, Aspergillus sp., Phomopsis sp., Stemphylium sp. were isolated from diseased root samples of date palm collected from different governorates. Also, eight different fungi, viz. Fusarium sp., Aspergillus sp., Thielaviopsis paradoxa, Mucor sp., Cladosporium sp., Stemphylium sp., Alternaria sp. and Rhizoctonia solani were isolated from rhizosphere samples of date palm collected from different governorates. Data in Table (2) show that diseased samples collected from Marsa-matrouh governorate gave four fungi isloated from roots, viz. F. oxysporum, F. solani, Phomopsis sp., and Stemphylium sp. (60, 30, 5 and 5%, respectively). Also, gave three fungi isolated from the rhizosphere, viz. F. solani, Aspergillus sp. and Rhizoctonia solani (70, 20 and 10%, respectively). Behaera governorate gave three fungi isolated from roots, viz. T. paradoxa, B. theobromae and F. semitectum (50, 30 and 20%, respectively). Also, gave five fungi isolated from the rhizosphere, viz. T. paradoxa, F. oxysporum,

Aspergillus sp., Cladosporium sp. and Mucor sp. (40, 20, 20, 10 and 10%, respectively). Sharkvia governorate gave two fungi isolated from roots, viz. F. oxysporum and F. semitectum (90 and 10%, respectively). Also, three fungi were isolated from the rhizosphere, viz. Rhizoctonia solani, Aspergillius sp. and F. semitectum (70, 20 and 10%, respectively). Ismailia governorate gave three fungi isolated from roots, viz. F. oxysporum, F. solani and Rhizoctonia solani (50, 35 and 15%, respectively). Also, four fungi were isolated from the rhizoshphere, viz. F. oxysporum, Rhizoctonia solani, Aspergillus sp. and Mucor sp. (70, 10, 10 and 10%, respectively). Giza governorate gave three fungi isolated from roots, viz. F. oxysporum, F. moniliforme and Gliocladium sp. (60, 30 and 10%, respectively). Also, four fungi were isolated from the rhizosphere, viz. Stemphylium sp., Aspergillus sp., Alternaria sp. and *Mucor* sp. (30, 30, 20 and 20%, respectively). Luxor governorate gave three fungi isolated from roots, viz. F. oxysporum, F. moniliforme and Aspergillus sp. (60, 25 and 15%, respectively). Also, two fungi were isolated from the rhizosphere, viz. F. semitectum and Aspergillius sp. (70 and 30%, respectively). Aswan governorate gave three fungi isolated from roots, viz. F. oxysporum, F. moniliforme and F. solani (60, 20 and 20%, respectively). Also, six fungi were isolated from the rhizosphere, viz. F. oxysporum, F. semitectum, F. solani, F. subglutinans, F. clamidospores and Aspergillus sp., being 30, 20, 20, 10, 10, and 10%, respectively. On the other hand, Marsa-Matrouh governorate showed the highest count of F. oxysporum (60%) from roots, while F. solani (70%) from the rhizosphere. Behera governorate showed the highest count of T. paradoxa (50 and 40%) from roots and rhizosphere, respectively, while B. theobromae showed 30% from roots. Sharkvia governorate showed the highest count of F. oxysporum (90%) from roots, while R. solani (70%) was from the rhizosphere. Ismailia governorate gave the highest count of F. oxysporum (50 and 70%) from roots and rhizosphere, respectively. while R. solani (15 and 10%) was obtained from roots and rhizosphere, respectively. Giza governorate samples gave the highest count of F. oxysporum and F. moniliforme (60 and 30%) from roots, respectively. Luxor governorate gave the highest count of F. oxysporum and F. moniliforme (60 and 25%) from roots, respectively. Aswan governorate yielded the highest count of F. oxysporum isolated from the roots (60%) and (30%) from the rhizosphere. In general, the highest fungus distribution in all governorates under study, was F. oxsporum, followed by T. paradoxa, while R. solani showed the lowest distibution in all governorates under study.

Table (2): Occurrence and frequency (%) of fungi isolated from decline root rot and soil of date palm in different governorates.

Motase-Mediculi         Sheary         10         Rocots         Financialista Sheary         60         60         80           Behirs         10         Ribitzonberre         Financialista Sheary         10	Governorates	Cultivar	Number of samples	Samples (Roots/Rhizosphere)	Associated fungi	Number of isolates	Frequency %
Siway         10         Roots         Funnopsis sp.         6         8           10         Rhizoshpere         Stemphyllum sp.         6         6         10           10         Rhizoshpere         Asperalim solvent         6         10         10           10         Rhizoshpere         Asperalim solvent         10         10         10           10         Rhizoshpere         Asperalim solvent         10         10         10           2aghloul         6         Roots         Asperalim solvent         20         10           2aghloul         6         Roots         Asperalim solvent         10         10           2aghloul         6         Roots         Asperalim solvent         10         10           2aghloul         6         Roots         Asperalim solvent         10         10           Asperalim solvent         Asperalim solvent         10         10         10         10           Asperalim solvent         Asperalim solvent         10         10         10         10         10         10         10         10         10         10         10         10         10         10         10         10         10					Fusarium oxysporum	09	09
Silvay         Total Plantochiper (Included Septembrillium Sp.)         Phonotopia Speciment (Included Septembrillium Sp.)         6         Rhizoshpere (Included Septembrillium Sp.)         Phonotopia Speciment (Included Septembrillium Sp.)         6         Rhizoshpere (Included Septembrillium Sp.)         Phonotopia Speciment (Included Septembrillium Sp.)         1016         Phonotopia Speciment (Included Septembrillium Sp.)         1017         Phonotopia Speciment (Included Septembrillium Sp.)         101         Phonotopia Speciment (Included Included Septembrillium Sp.)         101         Phonotopia Speciment (Included Included Includ			ę	Poofe	Fusarium solani	30	30
Silvey         Filtrochipere         Stemphyllium sp.         6         Recording sp.         106         Recording sp.         106         Recording sp.         106         Recording specified speedones         106         Recording speedones         106         106         Recording speedones         106			2	STOCK	Phomopsis sp.	2	5
Hayany	Marsa-Matronh	Siway			Stemphylium sp.	9	9
10   Rhizoshpere   Agergillius sp.   10   Rhizoshpere   Agergillius sp.   15   Rhizoshpere   Agergillius sp.   15   Rhizoshpere   Rizastium solarit moniliforme   15   Rhizoshpere   Rizastium moniliforme   15   Rhizoshpere   Rizastium solarit moniliforme   15   Rhizoshpere   Rizastium solarit moniliforme   15   Rhizoshpere   Rizastium moniliforme   15   Rhizoshper					Fusrium solani	105	20
Hayany			10	Rhizoshpere	Aspergillus sp.	30	20
Hayany					Rhizoctonia solani	15	10
Hayany         6         Roots         Finalization semilectume         20           Hayany         6         Rhizoshpere         Finalization semilectum         46         8           Zaghloul         6         Roots         Finalization semilectum         10         10           Samany         8         Rhizoshpere         Agaegillus sp.         23         10           Finalization semilectum         10         10         10         10           Rhizoshpere         Rhizoshpere         Finalization solution         10         10           Finalization solution         10         10         10         10           Finalization solution         10         10         10         10         10           Reprezionia solution         Finalization solution         11 <td></td> <td></td> <td></td> <td></td> <td>Thielaviopsis paradoxa</td> <td>50</td> <td>50</td>					Thielaviopsis paradoxa	50	50
Hayany         Rhizoshpere Tueserium sendozen Euserium oxygorum Samnany         FRizoshpere Tueserium oxygorum Sendozen Solari Euserium oxygorum Sendozen Solari Samnany         FRizoshpere Tueserium oxygorum Sendozen Solari Sendozen Sol			9	Roots	Botryodiplodia theobromae	30	30
Heyany         Entition of Entire Intervious					Fusarium semitectum	20	20
Figure   F	Rehera	Havany			Thielaviopsis paradoxa	92	40
Record					Fusarium oxysporum	46	20
Caghiou  6			9	Rhizoshpere	Aspergillus sp.	46	20
Samany					Cladosporium sp.	23	10
Caphiou  6   Roots   Fusarium soupsorum   10   126					Mucor sp.	23	10
Zaghloul         6         Rhizoshpere         Russitum senitectum         100           Samany         8         Roots         Fusarium senitectum         36           Samany         8         Rizoshpere         Fusarium sonitectum         36           Fusarium oxysporum         36         36           Rizoshpere         Rizoshpere         147           Rizoshpere         Rizoshpere         21           Aspergillus sp.         21           Mucor sp.         21           Medjhol         112           Roots         Fusarium oxysporum         60           Fusarium oxysporum         60           Alkernius sp.         26           Alkernius sp.         26           Alkernius sp.         26           Alkernium oxysporum         60           Fusarium monillionne         20           Alkernius sp.         26           Alkernius sp.         26           Alkernium sp.         20           Aspergillus sp.         20           Fusarium monillionne         20           Fusarium spletctum         20           Fusarium solgitutinans         20           Fusarium solgitutinans         20 <td></td> <td></td> <td>9</td> <td>Roots</td> <td>Fusarium oxysporum</td> <td>06</td> <td>06</td>			9	Roots	Fusarium oxysporum	06	06
Zaghloul         6         Rhizoshpere         Appendituits solemin         1456           Samany         8         Roots         Fusarium solemin         16           Samany         8         Rhizoshpere         Fusarium oxysporum         50           Rhizoshpere         Rhizoshpere         Rhizoshpere         15           Asperdituis solemin         21         21           Rhizoshpere         Rhizoshpere         Rhizoshpere         21           Medihol         8         Rhizoshpere         Reservation solemin         21           Alexandrum oxysporum         60         60           Medihol         12         Rhizoshpere         Asperdituits sp.         26           Medihol         12         Roots         Fusarium solenim         20           Residina soleni         20         Roots         Fusarium solenim         20 </td <td></td> <td>:</td> <td></td> <td></td> <td>Fusarium semitectum</td> <td>10</td> <td>10</td>		:			Fusarium semitectum	10	10
Record   Record   Residence   Residence	Sharkyia	Zaghloul			Rhizoctonia solani	126	70
Sameny         Roots         Flusarium oxysporum         18           Sameny         Rhizoshpere         Flustrium soleni         36           Sameny         Rhizoshpere         Rhizochorm         147           Rhizoshpere         Rhizoshpere         Rhizoshpere         21           Aspergillus sp.         20         39           Medjhol         12         Roots         Flusarium moniliforme         26           Medjhol         12         Roots         Flusarium moniliforme         26           Medjhol         12         Roots         Flusarium sp.         26           Medjhol         12         Rhizoshpere         Flusarium moniliforme         26           Medjhol         12         Rhizoshpere         Flusarium moniliforme         20           Medjhol         12         Rhizoshpere         Flusarium moniliforme         20           Medjhol         12         Roots         Flusarium moniliforme         20           Flusarium moniliforme         52         60           Flusarium moniliforme         52           Flusarium moniliforme         52           Flusarium moniliforme         52           Flusarium moniliforme         52			9	Rhizoshpere	Aspergillus sp	36	20
Samenty         Roots         Flushium solemi         50           Samenty         8         Rhizoshpere         Flushium solemi         16           Zaghloul         8         Rhizoshpere         Rayergillus sp. 10         21           Zaghloul         8         Rhizoshpere         Flushium oxysporum         60           Aspergillus sp. 10         10         30         10           Aspergillus sp. 10         20         15           Aspergillum sp. 10         15         15           Aspergillum sp. 10         16         175           Aspergillum sp. 10         16         175           Aspergillum sp. 10         176         176           Asper					Fusarium semitectum	18	10
Samany         Roots         Fixerium solani         35           Samany         8         Rhizoshpere         Rhizoctonia solani         147           Rhizoshpere         Roots         Fizsarium oxysporum         21           Zaghloul         8         Rhizoshpere         Fizsarium monillorme         30           Medjhol         12         Rhizoshpere         Aspergillus sp.         26           Medjhol         12         Roots         Fizsarium monillorme         26           Medjhol         12         Rhizoshpere         Fizsarium oxysporum         60           Medjhol         12         Rhizoshpere         Fizsarium oxysporum         60           Medjhol         12         Rhizoshpere         Fizsarium oxysporum         60           Fizsarium oxysporum         60         76         60           Fizsarium oxysporum         60         76           Fizsarium oxysporum         60         76           Fizsarium oxysporum         60         76           Fizsarium oxysporum         60         76           Fizsarium oxysporum         60         78           Fizsarium oxysporum         60         78           Fizsarium oxysporum         60					Fusarium oxysporum	50	50
Samany         Rhizoshpere Risasium oxysporum         Harboctonia solani (147)         147         148 <t< td=""><td></td><td></td><td>œ</td><td>Roots</td><td>Fusrium solani</td><td>35</td><td>35</td></t<>			œ	Roots	Fusrium solani	35	35
Samany         8         Rhizoshpere Roots         Fizsarium oxysporum Asperdillus sp. 121         147           Zaghloul         8         Roots         Fizsarium moniiflome Good Color (Color (Colo					Rhizoctonia solani	15	15
Redinoshpere         Rhizoshpere Asperglius sp. A	smailia	Samany			Fusarium oxysporum	147	0.2
Receipting Sp.   Aspergillus			α	Rhizoshnere	Rhizoctonia solani	21	10
Among Sport Telesarium moriliforme again moriliforme again moriliforme again moriliforme again moriliforme again statement again moriliforme again again again again again again again moriliforme again agai			,		Aspergillus sp.	21	10
Zaghloul         8         Roots         Fusarium novisporum of Glockaldum sp. (10 of Glockaldum sp. (11 of Glockaldum sp. (12 of Glockaldum sp. (12 of Glockaldum sp. (13 of Glockaldum sp. (14 of Glockaldum sp. (14 of Glockaldum sp. (15					Mucor sp.	21	10
Red   Roots   Russium moniliforme   30   10   10   10   10   10   10   10					Fusarium oxysporum	09	09
Zaghloul         8         Rhizoshpere Appergillus sp. 256         Cilicoladium sp. 39         10           Medjhol         12         Roots Fusarium sonjilforme Appergillus sp. 26         15         16           Medjhol         12         Rhizoshpere Appergillus sp. 25         15         16           Medjhol         20         Roots Fusarium sonjiforme Appergillus sp. 20         15         16           Medjhol         20         Roots Fusarium sonjiforme Appergillus sp. 20         16         17           Medjhol         20         Roots Fusarium solani monjiforme appergillum oxysporum appergillum oxysporum appergillum solani monjiforme appergillum			œ	Roots	Fusarium moniliforme	30	30
Zaghloul         8         Rhizoshpere Attennal sophyllum sp.         Stemphyllum sp.         39         Respecial sophyllum sp.         30         <					Gliocladium sp.	10	10
Medjhol         Relizoshpere         Aspergillus sp.	Giza	Zaghloul			Stemphylium sp.	39	30
Medjhol         12         Roots Rhizoshpere Fusarium sorjaniliforme         Fusarium sorjaniliforme         26           Medjhol         12         Roots Fusarium moniliforme         25         16           Medjhol         12         Rhizoshpere Fusarium semitectum         715         16           Medjhol         20         Roots Fusarium solarii moniliforme         20         20           Fusarium solarii moniliforme         20         78         178           Fusarium solarii moniliforme         20         178         178           Fusarium solarii moniliforme         52         178         178           Fusarium solarii moniliforme         26         178         178           Fusarium solarii moniliforme         26         178         178           Fusarium solarii moniliforme         26         178           Fusari			α	Rhizoshnere	Aspergillus sp.	39	30
Medjhol         12         Roots Fusarium moniliforme         Fusarium solitione         26           Medjhol         12         Rhizoshpere         Fusarium moniliforme         115           Medjhol         20         Roots         Fusarium solaritectum         175           Medjhol         20         Roots         Fusarium moniliforme         20           Fusarium solaritectum         60         20           Fusarium solaritectum         52           Fusarium solaritectum         62           Fusarium solaritectum         62           Fusarium solaritum solaritectum         62           Fusarium solaritum solaritectum         62           Fusarium solaritum solaritectum         62           Fusarium solaritum solaritectum         52           Fusarium solaritum solaritectum         26           Fusarium solaritum solaritectum         26           Fusarium solaritum solaritum solaritectum         26			,		Alternaria sp.	26	20
Medjhol         12         Roots         Fusarium oxysporum romiliforme popsporum romiliforme romi					Mucor sp.	26	20
Medjhol         12         Roots         Fusarium moniliforme         25           Medjhol         12         Rhizoshpere         Fusarium senitectum         175           Medjhol         20         Roots         Fusarium solaniforme         20           Fusarium solaniforme         20         Fusarium solaniforme         20           Fusarium solaniforme         62         Fusarium solaniforme         52           Fusarium solaniforme         62         Fusarium solaniforme         52           Fusarium solaniforme         52         Fusarium solaniforme         52           Fusarium clamidospores         26         Fusarium clamidospores         26					Fusarium oxysporum	09	09
Medjhol         12         Rhizoshpere Fusarlium soarlitectum         15         15           Medjhol         20         Roots         Fusarlium oxysporum         60         80           Medjhol         20         Rhizoshpere Fusarlium solari moniflorme         20         78           Fusarlium solari mostysporum         52         78           Fusarlium solari mostysporum         62         62           Fusarlium solari mostysporum         62         62           Fusarlium solari mostysporum         52         62           Fusarlium subglutinans         26         78           Fusarlium slappitus mostypores         26         78           Fusarlium slappitus mostypores         26         78			12	Roots	Fusarium moniliforme	25	25
12   Rhizoshpere   Fusarium semitectum   115   Fusarium Semitectum   115     20	Luxor	Medjhol			Aspergillus sp.	15	15
Medjhol         20         Roots         Fusarium solani         76           Medjhol         20         Rhizoshpere         Fusarium solani         20           Fusarium solani         Fusarium solani         78           Fusarium solani         62           Fusarium solani         62           Fusarium solani         52           Fusarium solani         52           Fusarium solani         52           Fusarium solani         52           Fusarium clanidospores         26           Fusarium clanidospores         26			12	Rhizoshpere	Fusarium semitectum	175	70
Medjhol         20         Roots         Fusarium novjsporum noniliforme         60           Medjhol         Fusarium solari mozysporum         20         78           Fusarium solari mozysporum         78         52           Fusarium solari mozysporum         62         52           Fusarium solari mozysporum         62         52           Fusarium subglutinans         26         76           Fusarium subglutinans         26         26           Aspergillus sp         26         Aspergillus sp			-:		Aspergillus sp.	75	30
20         Roots         Fusarium moniflorme         20           Fusarium solani         20         8           Medjhol         Fusarium semitectum         52           Fusarium solani         62           Fusarium solani         52           Fusarium subglutinans         26           Fusarium clamidospores         26           Aspergillus sp         26					Fusarium oxysporum	09	09
Medjhol         Eusarium solani         20           Rhizoshpere         Fusarium solani etum subgutunan subgutun subgutunan subgutunan subgutunan subgutunan subgutunan subgutun subgutu subgutu subgutun subgutun subgutun subgutu subgutun subgutu subgutun su			20	Roots	Fusarium moniliforme	20	20
Medjhol         Fusarium oxysporum         78           20         Rhizoshpere         Fusarium solarium solarium solarium subglutinans         52           Fusarium subglutinans         26           Fusarium clamidospores         26           Aspergillus sp         26					Fusarium solani	20	20
Medjhol         Fusarium semitectum         62           20         Rhizoshpere         Fusarium solani         52           Fusarium subglutinans         26         Fusarium clamidospores         26           Fusarium clamidospores         26         Aspergillus sp					Fusarium oxysporum	82	30
Flusarium solani	Aswan	Medjhol			Fusarium semitectum	52	20
Pusarium subglutinans			20	Rhizoshnere	Fusarium solani	52	20
26 26			3		Fusarium subglutinans	26	10
26					Fusarium clamidospores	26	10
					Aspergillus sp	26	10

# Pathogenicity test of the most frequent isolated fungi:

Different isolated fungi, viz. F. oxysporum, F. moniliforme, F. solani, F. semitectum, B. theobromae, T. paradoxa and R. solani from different locations and different governorates were used to investigate their pathogenic capabilities on young date palm seedlings, originated from seeds of three vars. Zaghloul, Sammany and Hayany. Data concerning the pathogenicity test on different wounded varieties of date palm seedlings after three months are shown in Table (3). It is evident that all tested fungi were able to induce root rot reaction, except F. semitectum. F. oxysporum isolate1 obtained from Aswan governorate was the most virulent one, where it showed 40.6% disease severity on the three varieties, followed by, F. oxysporum isolate 4 obtained from Luxor (31.8% DS). While, F. oxysporum isolates 2, 6, 3 and 5 gave 23.4, 20.1, 19.9 and 19.7% DS, respectively on the average. On the other hand, F. solani and T. paradoxa recorded 23.8 and 23.8% DS on the average. B. theobromae, F. moniliforme and R. solani recorded 19.5, 14.8 and 14.3%DS, respectively on the average for the three vars. Zaghloul variety was the most susceptible to all the tested fungi, except F. semitectum, followed by vars. Sammany and Hayany, respectively. Data also show that increased time after inoculation led to increasing the disease severity for all fungi and cultivars. Data in Table (4) indicate that using the unwounded method, F. oxysporum isolate1, obtained from Aswan governorate, was the most virulent one, 21.1% disease severity on the average for all three varieties, followed by, F. oxysporum isolate 4 obtained from Luxor (15.0%DS). While, F. oxysporum isolates 2, 5, 3 and 6 gave 10.5, 10.2, 9.9 and 9.1%DS, respectively. On the other hand, F. solani and T. paradoxa recorded 16.3 and 13.7%DS, on the average for the all three varieties. F. moniliforme, R. solani and B. theobromae recorded 10.2, 9.6 and 9.4%DS, respectively on the average. Zaghloul var. was the most susceptible with all the tested fungi, except F. semitectum, followed by vars. Sammany and Hayany, respectively. Data also show that increased the time after inoculation led to increasing the disease severity by all the tested fungi and varieties.

Table (3): Pathogenicity test using different varieties of wounded date palm seedlings after 30, 45, 60 and 90 days after inoculation.

							% Disease	% Disease severity on date palm vars.	on date	palmvars	٠					
Fungi			Zaghloul				<i>o,</i>	Sammany					Hayany			Mean
	30 days	45 days	45 days 60 days 90 days	90 days	Mean	30 days	45 days	30 days 45 days 60 days	90 days	Mean	30 days	30 days 45 days 60 days	60 days	90 days	Mean	mean
F. oxysporum1	37.5	54.2	58.3	70.8	55.2	25.4	33.3	39.2	42.1	35.0	22.9	26.3	33.3	44.2	31.7	40.6
F. oxysporum2	25.4	33.3	35.4	45.8	35.0	17.5	19.2	20.4	21.7	19.7	12.5	15.0	15.4	18.8	15.4	23.4
F. oxysporum3	22.9	31.3	37.5	37.5	32.3	10.4	13.3	15.4	17.9	14.3	8.3	12.5	13.3	17.9	13.0	19.9
F. oxysporum4	33.3	40.8	50.0	58.3	45.6	21.3	22.5	26.3	33.3	25.8	18.3	20.4	23.8	33.3	24.0	31.8
F. oxysporum5	25.0	35.4	37.5	39.2	34.3	0.0	13.3	14.2	14.2	10.4	11.7	13.3	16.3	16.3	14.4	19.7
F. oxysporum6	25.0	33.3	35.0	39.2	33.1	0.0	15.8	16.7	17.5	12.5	8.3	15.8	16.7	17.5	14.6	20.1
F. moniliforme	0.0	25.0	27.1	35.4	21.9	0.0	0.0	15.8	16.7	8.1	11.7	13.3	15.4	16.7	14.3	14.8
F. solani	0.0	28.1	37.5	46.9	28.1	18.3	18.8	23.1	23.1	20.8	18.3	21.3	23.1	27.5	22.6	23.8
F. semitectum	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
B. theobromae	0.0	29.2	33.3	38.3	25.2	15.8	17.5	19.2	20.4	18.2	7.5	13.3	17.9	21.3	15.0	19.5
I. Paradoxa	0.0	33.3	41.7	47.9	30.7	15.0	17.5	20.8	22.1	18.9	18.3	19.6	22.9	26.3	21.8	23.8
R. solani	0.0	25.0	29.5	31.3	21.4	0.0	9.2	13.3	14.2	9.2	8.3	10.8	14.2	15.8	12.3	14.3
Mean	14.1	30.7	35.2	40.9	30.2	10.3	15.0	18.7	20.3	16.1	12.2	15.1	17.7	21.3	16.6	21.0
L. S. D. at 0.05%	Fungi (F) Varieties (V) Period after F * V * T	Fungi (F) Varieties (V) Period after inoculation (T) F * V * T	oculatio	(E) u	2:2 1:1 1:3 7:6		> + + ×	3.8 4.4 2.2								

2.9

F \* 7 = 7 \* 4 × 7 = 7 \* 7 × 7 = 7 \* 7

1.5 0.9 5.1

Period after inoculation (T) F \* V \* T

Fungi (F) Varieties (V)

L. S. D. at 0.05%

Table (4): Pathogenicity test using different varieties of unwounded date palm seedlings after 30, 45, 60 and 90 days after inoculation.

							% Disease	% Disease severity on date palm vars.	on date	oalm vars	,;					
Fungi			Zaghloul				•	Sammany					Hayany			Mean
	30 days	45 days	45 days 60 days 90 days	90 days	Mean	30 days	45 days	30 days 45 days 60 days 90 days	90 days	Mean	30 days 45 days 60 days 90 days	45 days	60 days	90 days	Mean	mean
F. oxysporum1	0.0	28.8	33.3	37.5	24.9	13,3	19.2	20.8	23.8	19.3	13.3	19.2	20.8	23.8	19.3	21.1
F. oxysporum2	0.0	15.8	20.0	22.5	14.6	0.0	10.4	14.2	15.8	10.1	0.0	8.3	8.3	10.8	6.9	10.5
F. oxysporum3	0.0	13.3	19.2	20.8	13.3	0.0	8.3	12.5	14.6	8.9	0.0	8.3	10.4	11.7	9.7	9.9
F. oxysporum4	0.0	22.9	26.3	27.1	19.1	0.0	13.3	18.8	20.8	13.2	0.0	13.3	17.1	20.8	12.8	15.0
F. oxysporum5	0.0	18.3	21.7	23.3	15.8	0.0	0.0	13.3	14.2	6.9	0.0	8.3	10.8	12.5	6.7	10.2
F. oxysporum6	0.0	12.5	16.7	17.5	11.7	0.0	0.0	13.8	15.8	7.4	0.0	8.3	10.8	13.3	8.1	9.1
F. monili forme	0.0	13.3	15.8	20.0	12.3	0.0	8.3	15.4	17.5	10.3	0.0	8.3	10.8	12.5	7.9	10.2
F. solani	0.0	24.4	28.1	31.3	20.9	0.0	13.3	18.3	20.6	13.1	0.0	17.9	19.6	22.5	15.0	16.3
F. semitectum	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
B. theobromae	0.0	0.0	12.5	13.3	6.5	0.0	12.5	14.2	17.1	10.9	0.0	12.5	14.2	17.1	10.9	9.4
T. Paradoxa	0.0	0.0	22.5	25.4	12.0	0.0	15.4	19.2	22.9	14.4	0.0	16.3	19.2	23.3	14.7	13.7
R. solani	0.0	0.0	10.8	12.5	5.8	0.0	10.8	14.6	15.8	10.3	0.0	15.8	16.7	18.3	12.7	9.6
Mean	0.0	12.4	18.9	20.9	13.1	1.1	9.3	14.6	16.6	10.4	1.1	11.4	13.2	15.6	10.3	11.3

#### Varietal reaction:

Date palm offshoots of three varieties, viz. Zaghloul, Sammany and Hayany were used to study their susceptibility to the pathogenic fungi, F. oxysporum, F. moniliforme, F. solani, B. theobromae, T. paradoxa and R. solani. Data in Table (5) show that wounded offshoots of all the tested varieties were susceptible to infection at different levels. Zaghloul var. was the most susceptible with the tested fungi, followed by Hayany, while Sammany was the least susceptible. On the other hand, F. oxysporum and F. solani were the most virulent (23.01 and 14.35%DS, respectively), while T. paradoxa and F. moniliforme (12.59 and 10.46% DS, respectively) were moderately virulent, as well as B. theobromae and R. solani (9.63 and 8.24%DS, respectively) were the weak virulent. Increasing time after inoculation was significant with all fungi and cultivars. Data in Table (6) indicate that when unwounded method was used, all fungi were pathogenic to all varieties of date palm offshoots. F. oxysporum, F. moniliforme and T. paradoxa were the most virulent to date palm offshoots (13.33, 8.93 and 7.98%DS, respectively), while F. solani and B. theobromae (6.23 and 5.94%DS, respectively) were moderately virulent, while R. solani was the weak virulent (3.46%DS). On the other hand, Havany var. was the most susceptible to all pathogenic fungi, followed by Sammany which was moderately susceptible, while Zaghloul was the least susceptible one. Periods after inoculation were significant with all fungi and cultivars.

Table (5): Pathogenicity test on different cultivars of wounded date palm offshoots at 3, 6 and 9 months after inoculation.

				%Dis	eas e sevi	erity on d	%Dis eas e severity on date palm vars.	vars.					
Fungi		Zaghloul	loul			Sammany				Hayany			Main Mean
	3 Months	6 Months	9 Months	Mean	3 Months	3 6 Months Months	9 Months	Mean	3 Months	6 Months	9 Months	Mean	
F. oxysporum 1	20.42	24.17	45.42	30.00	15.83	16.67	21.67	18.06	18.75	20.83	23.33	20.97	23.01
F. solani	16.25	18.75	21.25	18.75	11.67	12.92	14.58	13.06	10.42	10.83	12.50	11.25	14.35
T. Paradoxa	13.33	15.42	17.50	15.42	9.58	11.25	13.33	11.39	10.42	10.83	11.67	10.97	12.59
F. moniliforme	10.42	11.67	15.42	12.50	7.08	8.75	10.83	8.89	7.50	10.42	12.08	10.00	10.46
B. theobromae	7.92	10.42	11.67	10.00	7.92	9.17	1125	9.45	8.33	8.75	11.25	9.44	69.6
R. sokni	7.50	8.33	10.83	8.89	5.42	5.83	8.33	6.53	8.33	9.17	10.42	9.31	8.24
Control	0.00	00'0	00.0	0.00	0.00	00.0	0.00	0.00	0.00	0.00	0.00	0.00	00.0
Mean	10.83	12.68	17.44	13.65	8.21	9.23	11.43	9.62	9.11	10.12	11.61	10.28	11.18
L. S. D. at 5%	Fungi (F) Cultivars (V) Period after F * V * T	Fungi (F) Cultivars (V) Period after inoculation (T) F * V * T	ılation (T		0.94 0.62 0.62 2.99			F * T	1.73 1.73 1.22				

Table (6): Pathogenicity test on different cultivars of unwounded date palm offshoots at 3, 6 and 9 months after inoculation.

					1								
					%Dis	ease s ev	%Disease severity on date palm vars.	ate pall	n vars.				
Fungi		Zaghloul	loul			Sammany	nany			Hayany	any		Main
	3 Months	6 Months	9 Months	Mean	3 Months	6 Months	9 Months	Mean	3 Months	6 Months	9 Months	Mean	Mean
F. oxysporum 1	12.50	14.17	17.08	14.58	11.67	15.83	18.33	15.28	7.92	10.42	12.08	10.14	13.33
T. Paradoxa	9.17	11.25	12.92	11.11	8.67	9.08	9.58	9.11	5.83	6.42	7.42	6.56	8.93
F. solani	6.25	7.50	8.75	7.50	7.08	8.33	11.67	9.03	5.83	7.92	8.50	7.42	7.98
F. moniliforme	3.67	6.33	6.83	5.61	3.50	6.25	8.33	6.03	5.33	7.08	8.75	7.05	6.23
B. theobromae	5.92	6.33	6.42	6.22	5.75	6.17	6.58	6.17	4.67	5.42	6.42	5.50	5.96
R. sokani	3.25	3.83	4.25	3.78	2.92	3.17	3.58	3.22	2.92	3.33	3.92	3.39	3.46
Control	00.00	00.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	00.0	0.00	0.00
Mean	5.82	7.06	8.04	6.97	99.6	86.9	8.30	86.9	4.64	5.80	6.73	5.72	95.9
L. S. D. at 5%					9			1					

0.73 0.73

0.43 0.28 0.28 n.s.

Fungi (F) Cultivars (V) Period after inoculation (T) F ^ V ^ T

#### **Discussion**

Date palms under the Egyptian conditions are subjected to infection with different diseases caused by many soil-borne pathogenic fungi causing considerable root rot in the orchards (Abbas et al., 1989 and El-Deep, 1994). The present study aimed to carry out an accurate: survey and identification of fungi associated with root rot of date palm. Many researchers reported that soil-borne pathogenic fungi are the mostly responsible for root rot diseases which cause considerable losses of date palm and offshoots (El- Deep, 1994 and Abdalla et al., 2000). They mentioned that F. oxysporum, F. solani and F. moniliforme were isolated from the declined date palm trees. Root rot caused by soil-borne pathogenic fungi, viz. F. oxysporum, F. moniliforme, F. solani, B. theomromae, T. paradoxa and R. solani were isolated from root rotted samples collected from Aswan. Behera. Giza, Ismailia, Luxor, Sharkvia and Marsa-Matrouh governorates. While other fungi were less frequent. Results of the present study indicated that the root rot diseases were noticed on date palm grown in different localities belonging to the seven governorates in Egypt. El-Arosi et al. (1983) reported that symptoms on date palm infected by F. moniliforme and F. solani appeared on root as pale brown discoloration on the adventitious roots. The occurrence and frequency of the isolated fungi were differed from one location to another; these differences are probably due to the environmental conditions such as moisture, temperature and soil type, dissemination factors of fungi in different locations and agricultural practices. These results are in harmony with those obtained by Fawcett and Klotz (1932). Variations were recorded on the disease incidence and disease severity percentage in the inspected governorates. These results are in agreement with those obtained by El-Deep et al. (2007) who mentioned that these variations might be due to the variation in environmental conditions. It may be also due to one or more of the following factors; 1- pathogen frequency, 2- climatic conditions which differ considerably between locations, 3- varietal sensitivity, 4dissemination factors available in the locality. 5- it may be also due to the cultural practices (Turner, 1981). Fusarium spp. were the most frequently isolated fungi from all governorates studied. Also, Fusarium spp. were the highly prevailed in healthy and infected date palm and offshoots. Mandel et al. (2005) showed that Fusarium solani and F. oxysporum were predominant in the rhizosphere of date palm. Fusarium spp. were the most frequently isolated fungi from roots, but they showed the highest mean recovery of Fusarium spp. from the plant debris followed by the roots and the lowest one was occurred in the soil samples. On the other hand, T. paradoxa was consistently isolated from rotted roots of date palm, these results are in agreement with those obtained by Al-Rokibah et al. (1998) and Samir et al. (2009). On the other hand, Dierbi (1991) mentioned that T. paradoxa was not isolated from the roots of naturally infested trees. This may be due to the presence of various biotypes of the fungus in different regions (Al-Rokibah et al., 1998). The pathogenic potentialities of the isolated fungi were determined with three date palm vars. In the greenhouse, all wounded seedlings of date palm varieties were infected with all soil-borne pathogenic fungi, but with various degrees of susceptibility. The most virulent fungus was Fusarium oxysporum, followed by T. paradoxa, while F. moniliforme, F. solani and B. theobromae were moderately virulent. R. soalni was weak virulent to date palm. Hayany variety was the most susceptible to root rot followed by Sammany var., while Zaghloul var. was the less susceptible. These results are in agreement with those obtained by El-Deep et al. (2007).

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# حصر وتعريف لأهم الفطريات التي تسبب مرض عفن الجذور وأهميتها على نخيل البلح في مصر

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تتعرض أشجار النخيل ، تحت الظروف المصرية للإصابة بالأمراض المختلفة التي تسبيها العديد من فطريات التربة مما يتسبب عنها تعفن جذور الشتلات والأشجار الكبيرة وموتها في حالات الإصابة الشديدة. أجرى الحصر المرضى خلال أربعة أعوام 2005-2008 لتحديد الفطريات التي تصيب جذور نخيل البلح في سبع محافظات (أسوان، الأقصر، مرسى مطروح ، الجيزة، الاسماعيلية، الشرقية والبحيرة). تم إختيار هذه المحافظات على أساس: المساحة المنزرعة، أصناف النخيل المختلفة، ونظام الري، والظروف الجوية المختلفة وإختلاف نوع التربة. وأظهرت النتائج أن أمراض أعفان الجذور وجدت في جميع المحافظات ولكن بنسب مختلفة من حيث نسبة وشدة الاصابة، والتي نتجت عن العديد من الفطريات الممرضة. وأوضحت نتائج التشخيص المرضى أن محافظة أسوان كانت أعلى في شدة الإصابة بنسبة (45.00٪)، تليها الأقصر (37.50٪)، البحيرة (30.50٪)، مرسى مطروح (25.00 ٪)، الاسماعيلية (5.00٪)، الشرقية (3.75 ٪)، والجيزة (2.50 ٪). وأظهرت نتائج العزل المعملي أن أعلى نسبة تكرار كانت للإنواع المختلفة لجنس فيوزاريوم، يليها الفطر ثيلافيبسوس بارادوكسا، وأخيرا الفطر بيترودبلوديا ثيوبرومي والفطر ريزوكتونيا سولاني. الفطر فيوزرايوم اوكسيسبورم كان الأشد في القدرة المرضية يليه الفطر فيوزاريوم مونيليفورم والفطر فيوزاريوم سولاني وأخيرا الفطر ثيلافيبسوس بارادوكسا وهي الفطريات المسئولة عن إحداث مرض عفن الجذور والتي تسبب الاصفرار التدريجي للاشجار المصابة ثم يليها موت الاشجار. جميع أصناف النخيل المختبرة كانت قابلة للإصابة بالفطريات المسببة لمر ض عفن الجذور . وكان الصنف حياني الأكثر قابلية للإصابة يليه الصنف سماني و أخير ا الصنف زغلول.